

Dorota SZAL<sup>1</sup>  
Renata GRUCA-ROKOSZ<sup>2</sup>

## ANAEROBIC OXIDATION OF METHANE IN FRESHWATER ECOSYSTEMS

Anaerobic oxidation of methane (AOM) is a biochemical process that plays an important role in aquatic ecosystems, as it significantly reduces the emission of methane (CH<sub>4</sub>) to the atmosphere. Under anaerobic conditions, CH<sub>4</sub> can be oxidized with electron acceptors, such as sulphates (SO<sub>4</sub><sup>2-</sup>), nitrates (NO<sub>3</sub><sup>-</sup>) or nitrites (NO<sub>2</sub><sup>-</sup>), iron (Fe<sup>3+</sup>), manganese (Mn<sup>4+</sup>) and humic substances. The anaerobic oxidation of methane is mainly regulated by anaerobic methanotrophic archaea (ANME) and sulphate reducing bacteria. The AOM process is crucial to understand the CH<sub>4</sub> cycle and anticipate future emissions of the gas from water reservoirs. The process is widely described in marine environments, however very little is known about its occurrence and importance in freshwater systems. There is a great demand for this kind of the research, especially in ecosystems exposed to long-term anaerobic conditions, which may be in degraded reservoirs.

**Keywords:** anaerobic oxidation of methane, electron acceptors, methanotrophic archaea

### 1. Introduction

Methane (CH<sub>4</sub>) is a gas emitted to the atmosphere from both natural and anthropogenic sources. It contributes to the greenhouse effect and global climate change [24]. Production of CH<sub>4</sub> occurs under anaerobic conditions with the participation of methanogenic archaea; in turn, methane oxidizing bacteria (methanotrophic archaea) contribute to reducing the emission of this gas to the atmosphere. Although aerobic methane-oxidizing bacteria (MOB) have been known for over 100 years [60], it was only at the turn of the last century that organisms involved in anaerobic oxidation of methane (AOM) were identified. Anaerobic methanotrophic archaea (ANME) are of great importance in regulating the Earth's climate, because they reduce the emission of large

---

<sup>1</sup> Corresponding author: Dorota Szal, Politechnika Rzeszowska, ul. Powstańców Warszawy 6, 35-959 Rzeszów, d.piwinska@prz.edu.pl

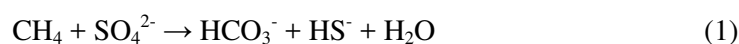
<sup>2</sup> Renata Gruca-Rokosz, Politechnika Rzeszowska, ul. Powstańców Warszawy 6, 35-959 Rzeszów, renatagr@prz.edu.pl

amounts of CH<sub>4</sub> accumulated in bottom sediments [32]. The process of anaerobic oxidation of methane (AOM) is widely described in marine environments, however very little is known about its occurrence and importance in freshwater systems [8; 26]. Just a decade ago, sulfates were the only known electron acceptor supporting anaerobic oxidation of methane [32; 38]; however, there are many other electron acceptors in the environment, such as: iron (Fe<sup>3+</sup>), manganese (Mn<sup>4+</sup>), nitrates (NO<sub>3</sub><sup>-</sup>) [19] or nitrites (NO<sub>2</sub><sup>-</sup>) [14; 48], which are thermodynamically more preferred than sulfates.

## 2. Anaerobic oxidation of methane

Methane emission from sediments is the net result of two processes: methanogenesis, which occurs in the hypoxic part of bottom sediments, and the oxidation of CH<sub>4</sub> as a result of aerobic or anaerobic microbiological processes [42]. AOM is an important process in marine and freshwater ecosystems [33; 44; 45], because it plays a key role in reducing the flow of CH<sub>4</sub> from bottom sediments to the overlying water, and thus also to the atmosphere [5; 34; 37]. AOM mainly occurs in the presence of methanotrophic archaea and sulfate-reducing bacteria [7]. AOM plays an important role in controlling CH<sub>4</sub> emissions in marine sediments, where it is oxidized on average 300–380 Tg CH<sub>4</sub>·yr<sup>-1</sup> [20; 46]. It has also been estimated that, for example, in wetlands the AOM process can consume 4.1-6.1 Tg CH<sub>4</sub>/m<sup>2</sup>·yr, which is about 2-6% of the CH<sub>4</sub> emissions from wetlands in the world [23]. In the absence of oxygen, microorganisms can oxidize CH<sub>4</sub> in the presence of alternative electron acceptors, such as: sulfates, nitrates, nitrites, iron, manganese and humic substances [6; 7; 9; 14; 15; 18; 19; 21; 39; 45; 48; 49; 54; 56; 63].

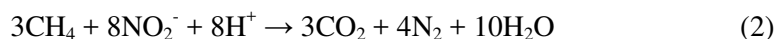
Anaerobic oxidation of methane coupled with sulfate reduction (AOM-SR) is the main process contributing to a significant reduction of methane produced in marine sediments. AOM-SR is regulated by a consortium of anaerobic methanotrophic archaea (ANME) and sulfate-reducing bacteria [32]. Methane is oxidized according to reaction:



The AOM-SR process is influenced by the distribution of CH<sub>4</sub> and SO<sub>4</sub><sup>2-</sup> in bottom sediments [26], as well as the stable isotopic composition δ<sup>13</sup>C-CH<sub>4</sub> [1]. There is little evidence of the role of this process in freshwater sediments, where the reduction of sulfate is limited. However, the results of the research presented in some publications [42; 51] indicate that AOM-SR can play a significant role in regulating the flow of CH<sub>4</sub> from sediments for SO<sub>4</sub><sup>2-</sup> concentrations typical for freshwater reservoirs. AOM dependent from SO<sub>4</sub><sup>2-</sup> concentrations have been demonstrated in the bottom sediments of Lake Cadagno (Switzerland) based on the isotopic analysis of carbon and archaea involved in the AOM. This process

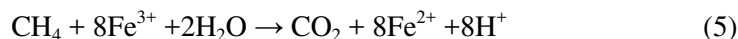
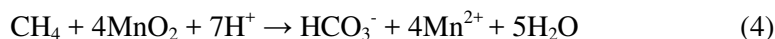
was particularly present near the surface of the sediment, where the concentration of  $\text{SO}_4^{2-}$  was  $> 2$  mmol/l [52]. In turn, based on the research of some authors [42; 51], it was concluded that in a monomictic lake in Denmark (lake Ørn) high concentrations of sulfates may adversely affect the thermodynamics of the AOM process. Therefore, further research is needed to quantify the role of AOM in the  $\text{CH}_4$  cycle in various freshwater environments.

Electron acceptors, i.e. nitrates, nitrites, iron and manganese, may play a potentially greater role in freshwater ecosystems [42]. Due to the fact that  $\text{SO}_4^{2-}$  concentrations are generally at low level in freshwater lakes,  $\text{NO}_3^-$  and  $\text{NO}_2^-$  may play a greater role in the AOM process. If nitrates and/or nitrites are available in anaerobic sediments, anaerobic bacteria oxidizing nitrates and/or nitrites take part in the AOM process. In this case,  $\text{CH}_4$  can be used as an electron donor (reactions 2 and 3) [62], and sulfate reduction usually does not occur [50]. This process is called anaerobic oxidation of methane coupled with denitrification (AOM-D) [11; 41].



The AOM-D process is thermodynamically more favorable than AOM-SR. Therefore, AOM-D has become the subject of many scientific studies [14; 25; 45]. AOM-D was observed in lake sediments with a high concentration of  $\text{NO}_3^-$  [12; 37].

It is also possible to oxidize  $\text{CH}_4$  by reducing manganese (Mn) or iron (Fe) (reactions 4 and 5) as confirmed by recent studies [6; 10; 53].



In the environment, iron reduction is often limited by the bioavailability of iron oxides, as iron (III) or complex iron compounds are not commonly found. In the publication of Ettwig et al. [17], the enrichment culture was incubated with more available metal forms: nanoparticle ferrihydrite ( $\text{Fe}^{3+}$ ) and birnessite ( $\text{Mn}^{4+}$ ). Selected electron acceptors supported the oxidation of  $\text{CH}_4$  to  $\text{CO}_2$ , although the process itself was at a lower rate compared to nitrates and Fe (III) as iron citrate as electron acceptors. Stimulation of AOM after the addition of oxidized forms of Mn or Fe was observed in laboratory incubations of  $\text{SO}_4^{2-}$ -depleted freshwater sediments. It was found that iron reduction affects AOM in lake sediments, as indicated by higher  $\delta^{13}\text{C}-\text{CH}_4$  values in deeper sediment layers [53]. AOM coupled with reduction of Mn and/or Fe was also observed in the water column of thermally stratified Lake Matano (Indonesia), the world's largest known ferruginous reservoir [10]. These studies have shown that the role

of Mn and Fe oxides in AOM is indirect and involves the stimulation of oxidative sulfur circulation, which can supply  $\text{SO}_4^{2-}$  to AOM-SR. Based on the study of lake Ørn sediments (Denmark), it was concluded that the process took place at significant depths, where both iron and sulfates were present in low concentrations, confirming the significance of AOM-Fe/Mn in Fe/Mn-depleted sediments. Furthermore, anaerobic reduction of iron ( $\text{Fe}^{+3}$ ) and manganese ( $\text{Mn}^{+4}$ ) coupled with AOM is regarded as more favorable process; and it has been shown that it is a significant process in lakes [42; 53]. Little is known about the microorganisms involved in AOM-Fe/Mn. However, on the basis of the conducted research [6], it was noticed that a large group of microorganisms of AOM-Fe/Mn in sediments after incubation were microorganisms associated with the marine benthic Archaea group D and ANME [42].

The rate of anaerobic oxidation of  $\text{CH}_4$  depends not only on the availability and concentration of anaerobic oxidants, but also on the weather conditions and physical factors prevailing on the reservoir, such as water turbulence [3; 31]. High wind speeds can lead to complete mixing of the reservoir waters in a short time, e.g. in autumn. Therefore, the majority of  $\text{CH}_4$  can be emitted to the atmosphere due to increased transport through different water layers and a short residence time in the water column [3; 29]; on the other hand, gradual mixing of reservoir water may cause that  $\text{CH}_4$  will be longer available for bacteria participating in AOM [29], which will reduce its emission to the atmosphere.

### 3. Anaerobic methanotrophic archaea

Anaerobic methanotrophic archaea (ANME) are involved in anaerobic methane oxidation – in the process opposite to methanogenesis [32]. Based on the phylogenetic analysis of the 16S rRNA gene, ANME were divided into three groups: ANME-1, ANME-2 and ANME-3 [8; 21; 33; 40]. All ANMEs are phylogenetically linked to various groups of methanogenic archaea. ANME-2 and ANME-3 belong to the order *Methanosarcinales*; in turn, ANME-1 belong to the order related to *Methanosarcinales* and *Methanomicrobiales* [32]. ANME-3 are closely related to the *Methanococcoides* gene. Microorganisms belonging to the ANME-2 and ANME-3 groups have a shape similar to methanogens of *Methanosarcina* and *Methanococcus*; whereas ANME-1 show a different morphology. ANME strains occur in anaerobic freshwater sediments, as well as in marine environments, aquifers and soils [32; 58].

AOM-SR is a metabolic process combined with  $\text{SO}_4^{2-}$  reduction, obtaining energy through syntrophic consortium of the ANME and sulfate reducing bacteria (SRB) [7; 22]. Some SRB groups: *Desulfosarcina/Desulfococcus* and *Desulfobulbaceae* participate in the sulfate reduction (SR) process together with ANME. However, it has been shown that some ANME filotypes perform SR without the presence of SRB in various aquatic environments [2; 35; 36; 59; 61], suggesting that AOM-SR may occur regardless of the occurrence of ANME [38].

In addition, Ettwig et al. [14] showed that AOM can be mediated via *Candidatus Methyloirabilis oxyfera* (NC10 type) in the presence of  $\text{NO}_2^-$ . The bacterium can produce oxygen by reducing  $\text{NO}_2^-$  and use the  $\text{O}_2$  to oxidize  $\text{CH}_4$  [11; 14]. *Methyloirabilis oxyfera*, an NC10 bacterium, and *Candidatus Methanoperedens nitroreducens*, ANME archaea, have been identified as microorganisms capable of carrying out AOM-D [14; 19; 37]. Denitrification metanotrophs seem to be common in freshwater sediments, as determined by 16S rDNA studies [16], but the quantitative role of  $\text{NO}_3^-$  or  $\text{NO}_2^-$  in AOM under natural conditions has not been studied so far.

AOM associated with the reduction of soluble iron complexes have recently been observed in environments rich in ANME-2 [49], and AOM coupled with reduction of iron or manganese oxides has been confirmed several times and presented in many scientific publications [6; 13; 43; 47; 53]. In addition to the ability of these bacteria to carry out AOM along with iron reduction in enrichment cultures [17], it was found that ANME-2d or AAA (*AOM-associated archaea*) containing *Candidatus Methanoperedens nitroreducens* carry out AOM-SR in the sediments of the alpine lake Cadagno (Switzerland) [52; 57]. Recent studies have concluded that the strain closely related to *Candidatus Methanoperedens nitroreducens* from freshwater enrichment culture is present during AOM coupled with the reduction of soluble and nanoparticulate iron forms [17].

Methane oxidizing bacteria not only reduce atmospheric  $\text{CH}_4$  emissions, but also provide adequate nutrients for aquatic consumers [27]. In several scientific publications [4; 28; 30; 55] the role of  $\text{CH}_4$  oxidizing bacteria was examined as a carbon source for zooplankton in humic lakes with thermal stratification of water. Kankaala et al. [30] showed that metanotrophs were a source of nutrients for a typical pelagic zooplankton – *Daphnia*. The authors also suggested that carbon from  $\text{CH}_4$  plays a greater role in the trophic network of lakes than it was previously estimated.

#### 4. Summary

Freshwater ecosystems are identified as one of the main natural sources of methane, but little is known about the importance of anaerobic methane oxidation (AOM) in these ecosystems. The emerging publications of many authors show that AOM can significantly reduce methane emissions from freshwater sediments. The study of the activity of microorganisms taking part in AOM and the ongoing metabolic processes with the participation of electron acceptors ( $\text{Fe}^{3+}$ ,  $\text{Mn}^{4+}$ ,  $\text{SO}_4^{2-}$ ,  $\text{NO}_3^-$ ,  $\text{NO}_2^-$ ) is crucial in understanding the circulation and predicting future  $\text{CH}_4$  emissions. The flow of methane into the atmosphere from aquatic ecosystems is regulated by two main groups of microorganisms. Methanogenic archaea are responsible for the production of  $\text{CH}_4$ , while methane-oxidizing bacteria (ANME) are responsible for the

consumption of CH<sub>4</sub> in these ecosystems. Different electron acceptors contained in sediments affect the presence of various groups and species of microorganisms characteristic of individual processes: AOM-SR, AOM-D and AOM-Fe/Mn. In marine environments, the dominant process is AOM coupled with sulfate reduction. In freshwater ecosystems, this process is probably limited by low sulfate concentrations. The methane oxidation processes associated with the reduction of alternative electron acceptors such as: nitrates/nitrites, manganese and iron are more significant there.

*The work was supported by National Science Centre Poland, via grant no. 2017/25/B/ST10/00981.*

## References

- [1] Alperin M.J., Reeburgh W.S., Whiticar M.J., Carbon and hydrogen isotope fractionation resulting from anaerobic methane oxidation, *Global Biogeochem. Cycles*, 2, 1988, 279–288.
- [2] Aquilina A., Knab N.J., Knittel K., Kaur G., Geissler A., Kelly S.P., Fossing H., Boot C.S., Parkes R.J., Mills R.A., Boetius A., Lloyd J.R., Pancost R.D., Biomarker indicators for anaerobic oxidizers of methane in brackish-marine sediments with diffusive methane fluxes, *Organic Geochemistry* 41, 2010, 414–426.
- [3] Bastviken D., Cole J.J., Michael L. Pace Matthew C. Van de Bogert, Fates of methane from different lake habitats: Connecting whole-lake budgets and CH<sub>4</sub> emissions, *Journal of geophysical research*, 113, 2008, doi:10.1029/2007JG000608.
- [4] Bastviken, D., Ejlertsson, J., Sundh, I., Tranvik, L., Methane as a source of carbon and energy for lake pelagic food webs, *Ecology* 84, 2003, 969–981.
- [5] Bastviken D., Ejlertsson J., and Tranvik L., Measurement of methane oxidation in lakes—A comparison of methods, *Environ. Sci. Technol.*, 36, 2002, 3354–3361.
- [6] Beal E.J., House C.H., Orphan V.J., Manganese- and Iron-Dependent Marine Methane Oxidation, 325(5937), 2009, 184–187, DOI: 10.1126/science.1169984.
- [7] Boetius A., Ferdelman T., Lochtea K., Bacterial activity in sediments of the deep Arabian Sea in relation to vertical flux, *Deep Sea Research Part II: Topical Studies in Oceanography*, 47(14), 2000a, 2835–2875.
- [8] Boetius A., Ravensschlag K., Schubert C. J., Rickert D., Widdel F., Gieseke A., Amann R., Jørgensen B. B., Witte U., Pfannkuche O., A marine microbial consortium apparently mediating anaerobic oxidation of methane, *Nature*, 407, 2000b, 623–626, doi:10.1038/35036572.
- [9] Caldwell S.L., Laidler J.R., Brewer E.A., Eberly J.O., Sandborgh S.C., Colwell F.S., Anaerobic oxidation of methane: Mechanisms, bioenergetics, and the ecology of associated microorganisms, *Environ Sci Technol.* 42, 2008, 6791–6799.
- [10] Crowe S.A., Katsev S., Leslie K., et al., The methane cycle in ferruginous Lake Matano, *Geobiology* 9(1), 2011, 61–78.
- [11] Deutzmann J.S., Hoppert M., Schink B., Characterization and phylogeny of a novel methanotroph, *Methyloglobulus morosus* gen. nov., spec. nov. *Sys. Appl. Microbiol.* 37, 2014, 165–169, doi: 10.1016/j.syapm.2014.02.001.

- [12] Deutzmann, J. S., Schink B., Anaerobic Oxidation of Methane in Sediments of Lake Constance, an Oligotrophic Freshwater Lake, *Applied and Environmental Microbiology*, 77, 2011, 4429–4436.
- [13] Egger M., Jilbert T., Behrends T., Rivard C., Slomp C. P., Vivianite is a major sink for phosphorus in methanogenic coastal surface sediments, *Geochimica et Cosmochimica Acta*, 169, 2015, 217–235.
- [14] Ettwig K.F., Butler M.K., Le Paslier D. et al., Nitrite-driven anaerobic methane oxidation by oxygenic bacteria, *Nature* 464(7288), 2010, 543–548.
- [15] Ettwig K.F., Shima S., van de Pas-Schoonen K.T., Kahnt J., Medema M.H., op den Camp H.J.M., Jetten M.S.M., Strous M., Denitrifying bacteria anaerobically oxidize methane in the absence of Archaea, *Environmental Microbiology*, 10(11), 2008, 3164–3173, doi:10.1111/j.1462-2920.2008.01724.x.
- [16] Ettwig K.F., van Alen T., van de Pas-Schoonen K.T., Jetten M.S.M., Strous M., Enrichment and molecular detection of denitrifying methanotrophic bacteria of the NC10 phylum, *Appl. Environ. Microbiol.*, 75, 2009, 3656–3662.
- [17] Ettwig K., Zhu B., Speth D., Keltjens J.T., Jetten M.S.M., Kartal B., Archaea catalyze iron-dependent anaerobic oxidation of methane, *PNAS*, 113, 45, 2016, 12792–12796, [www.pnas.org/cgi/doi/10.1073/pnas.1609534113](http://www.pnas.org/cgi/doi/10.1073/pnas.1609534113).
- [18] Harder, J., Anaerobic methane oxidation by bacteria employing (super 14) C-methane uncontaminated with (super 14) C-carbon monoxide, In T. C. E. van Weering, G. T. Klaver, and R. A. Prins (ed.), *Marine geology*, Elsevier, Amsterdam, The Netherlands, vol. 137, 1997, 13–23.
- [19] Haroon MF, et al., Anaerobic oxidation of methane coupled to nitrate reduction in a novel archaeal lineage, *Nature* 500(7464), 2013, 567–570.
- [20] Hinrichs K.-U., Boetius A., The Anaerobic Oxidation of Methane: New Insights in Microbial Ecology and Biogeochemistry, *Ocean Margin Systems*, 2002, 457–477.
- [21] Hinrichs K.-U., Hayes J.M., Sylva S.P., Brewer P.G., DeLong E.F., Methane-consuming archaeobacteria in marine sediments, *Nature*, 398, 1999, 802–805.
- [22] Hoehler T., M.J. Alperin, D.B. Albert, C.S. Martens, Field and laboratory studies of methane oxidation in an anoxic marine sediment: Evidence for a methanogen-sulfate reducer consortium, *Global Biogeochemical Cycles* 8(4), 1994, 451–463.
- [23] Hu B., Shen L., Lian X., Zhu Q., Liu S., Huang Q., He Z., Geng S., Cheng D., Lou L., Xu X., Zheng P., He Y., Evidence for nitrite-dependent anaerobic methane oxidation as a previously overlooked microbial methane sink in wetlands, *PNAS* 111(22), 2014, 4495–4500.
- [24] Intergovernmental Panel on Climate Change, *Climate Change 2014, Mitigation of Climate Change, Summary for Policymakers and Technical Summary*, 2015.
- [25] Islas-Lima S., Thalasso, F., Gomez-Hernandez, J., Evidence of anoxic methane oxidation coupled to denitrification, *Water Res.*, 38, 2004, 13–16.
- [26] Iversen N., Jørgensen B.B., Anaerobic methane oxidation rates at the sulfate-methane transition in marine sediments from Kattegat and Skagerrak (Denmark), *Limnology and Oceanography* 30(5), 1985, 944–955, DOI: 10.4319/lo.1985.30.5.0944.
- [27] Jones, R. I., Grey, J., Biogenic methane in freshwater food webs, *Freshw. Biol.* 56, 2011, 213–229. doi: 10.1111/j.1365-2427.2010.02494.x.
- [28] Jones R.L.I., Whatley R.C., Cronin T.M., Dowsett H.J., Reconstructing late Quaternary deep-water masses in the eastern Arctic Ocean using benthonic ostracoda,

- Marine Micropaleontology, 37(3–4), 1999, 251–272, [https://doi.org/10.1016/S0377-8398\(99\)00022-5](https://doi.org/10.1016/S0377-8398(99)00022-5).
- [29] Kankaala, P., Eller G., Jones R.I., Could bacterivorous zooplankton affect lake pelagic methanotrophic activity, *Fundamental and Applied Limnology*, 169, 2007a, 203–209.
- [30] Kankaala P., Taipale S., Grey J., Sonninen E., Arvola L., Jones R.I., Experimental  $\delta^{13}\text{C}$  evidence for a contribution of methane to pelagic food webs in lakes, *Limnol. Oceanogr.* 51(6), 2006, 2821–2827.
- [31] Kankaala, P., Taipale S., Nykänen H., Jones R. I., Oxidation, efflux and isotopic fractionation of methane during autumnal turnover in a polyhumic, boreal lake, *Journal of Geophysical Research–Biogeosciences*, 112, 2007b, doi: 10.1029/2006JG000336.
- [32] Knittel K., Boetius A., Anaerobic oxidation of methane: Progress with an unknown process. *Annual Reviews of Microbiology*, 63, 2009, 311–334.
- [33] Knittel K., Lösekann T., Boetius A., Kort R., Amann R., Diversity and Distribution of Methanotrophic Archaea at Cold Seeps, *Applied and Environmental Microbiology*, 71(1), 2005, 467–479, <https://doi.org/10.1128/AEM.71.1.467-479.2005>.
- [34] Lofton D., Whalen S. C., Hershey A. E., Effect of temperature on methane dynamics and evaluation of methane oxidation kinetics in shallow Arctic Alaskan lakes, *Hydrobiologia* 721(1), 2014, DOI: 10.1007/s10750-013-1663-x.
- [35] Lösekann T., Knittel K., Nadalig T., Fuchs B., Niemann H., Boetius A. et al., Diversity and abundance of aerobic and anaerobic methane oxidizers at the Haakon Mosby mud volcano, Barents Sea, *Appl. Environ. Microbiol.*, 73, 2007, 3348–3362.
- [36] Maignien L., Parkes R.J., Cragg B., Niemann H., Knittel K., Coulon S., Akhmetzhanov A., Boon N., Anaerobic oxidation of methane in hypersaline cold seep sediments, *FEMS Microbiol. Ecol.*, 83, 2013, 214–231.
- [37] Martinez-Cruz K., Leewis M.-C., Herriott I. C., Sepulveda-Jauregui A., Anthony K. W., Thalasso F., Leigh M. B., Anaerobic oxidation of methane by aerobic methanotrophs in sub-Arctic, *Science of the Total Environment*, 607–608, 2017, 23–31.
- [38] Milucka J., et al., Zero-valent sulphur is a key intermediate in marine methane oxidation. *Nature*, 491(7425), 2012, 541–546.
- [39] Moran J.J., House C.H., Freeman K.H., Ferry J.G., Trace methane oxidation studied in several Euryarchaeota under diverse conditions, *Archaea*, 1, 2005, 303–309.
- [40] Niemann H., Duarte J., Hensen C., Omorigie E., Magalhaes V.H., Elvert M., Pinheiro L.M., Kopf A., Boetius A., Microbial methane turnover at mud volcanoes of the Gulf of Cadiz, *Geochimica et Cosmochimica Acta*, 70, 2006, 5336–5355.
- [41] Nordi K.A., Thamdrup B., Nitrate-dependent anaerobic methane oxidation in a freshwater sediment, *Geochim Cosmochim Acta*, 132, 2014, 141–150.
- [42] Nordi K., Thamdrup B., Schubert C. J., Anaerobic oxidation of methane in an iron-rich Danish freshwater Lake sediment, *Limnol. Oceanogr.*, 58(2), 2013, 546–554, doi:10.4319/lo.2013.58.2.0546.
- [43] Oni O., Miyatake T., Kasten S., Richter-Heitmann T., Fischer D., Wagenknecht L., et al., Distinct microbial populations are tightly linked to the profile of dissolved iron in the methanic sediments of the Helgoland mud area, North Sea. *Front. Microbiol.*, 6(365), 2015, doi: 10.3389/fmicb.2015.00365.
- [44] Pancost R.D., Sinninghe Damsté J.S., de Lint S., van der Maarel M.J., Gottschal J.C., Biomarker evidence for widespread anaerobic methane oxidation in Mediterranean sediments by a consortium of methanogenic archaea and bacteria, *Applied and Environmental Microbiology*, 66, 2000, 1126–1132.



- [45] Raghoebarsing A.A., Pol A., van de Pas-Schoonen K.T., Smolders A.J., Ettwig K.F., Rijpstra W.I., Schouten S., Damsté J.S., Op den Camp H.J., Jetten M.S., Strous M., A microbial consortium couples anaerobic methane oxidation to denitrification, *Nature*, 440(7086), 2006, 918–921.
- [46] Reeburgh, W., Oceanic methane biogeochemistry, *Chem. Rev.*, 107, 2007, 486–513.
- [47] Riedinger N., Formolo M. J., Lyons T. W., Henkel S., Beck A., Kasten S., An inorganic geochemical argument for coupled anaerobic oxidation of methane and iron reduction in marine sediments, *Geobiology*, 12(2), 2014, 172–181.
- [48] Roland F. A. E., Morana C., Darchambeau F., Crowe S. A., Thamdrup B., Descy J.-P., Borges A. V., Anaerobic methane oxidation and aerobic methane production in an east African great lake (Lake Kivu), *Journal of Great Lakes Research*, 2018, (in press).
- [49] Scheller S., Yu H., Chadwick G. L., McGlynn S. E., Orphan V. J., Artificial electron acceptors decouple archaeal methane oxidation from sulfate reduction, *Science*, 351(6274), 2016, 703–707, DOI: 10.1126/science.aad7154.
- [50] Schlesinger W.H., Bernhardt E.S., *Biogeochemistry: an analysis of global change*, 2013.
- [51] Schubert C. J., et al., Oxidation and emission of methane in a monomictic lake (Rotsee, Switzerland), *Aquat. Sci.*, 72, 2010, 455–466, doi:10.1007/s00027-010-0148-5.
- [52] Schubert C.J., Vazquez F., Lösekann-Behrens T., Knittel K., Tonolla M., Boetius A., Evidence for anaerobic oxidation of methane in sediments of a freshwater system (Lago di Cadagno), *FEMS Microbiol Ecol.*, 76, 2011, 26–38.
- [53] Sivan O., Adler M., Pearson A. et al., Geochemical evidence for iron mediated anaerobic oxidation of methane, *Limnol Oceanogr.*, 56, 2011, 1536–1544.
- [54] Smemo K.A., Yavitt J.B., Anaerobic oxidation of methane: an underappreciated aspect of methane cycling in peatland ecosystems?, *Biogeosciences*, 8, 2011, 779–793, <https://doi.org/10.5194/bg-8-779-2011>.
- [55] Taipale, S., Kankaala P., Jones R. I., Contributions of different organic carbon sources to *Daphnia* in the pelagic food web of a small polyhumic lake: Results from mesocosm <sup>13</sup>C-additions, *Ecosystems (N. Y., Print)*, 2008, doi:10.1007/s10021-007-9056-5.
- [56] Thauer R.K., Shima S., Methane as fuel for anaerobic microorganisms, *Ann N Y Acad Sci.*, 1125, 2008, 158–170.
- [57] Timmers P.H., Suarez-Zuluaga D.A., van Rossem M. et al., Anaerobic oxidation of methane associated with sulfate reduction in a natural freshwater gas source, *ISME J*, 10, 2016, 1400–1412.
- [58] Timmers P.H., Welte C.U., Koehorst J.J., et al., Reverse methanogenesis and respiration in methanotrophic Archaea, *Archaea*, 2017.
- [59] Treude T., Knittel K., Blumenberg M., Seifert R., Boetius A., Subsurface microbial methanotrophic mats in the Black Sea, *Appl. Environ. Microb.*, 71(10), 2005, 6375–6378.
- [60] Trotsenko Y.A., Murrell J.C., Metabolic aspects of aerobic obligate methanotrophy, *Advances in Applied Microbiology*, 63, 2008, 183–229.
- [61] Wankel S.D., Adams M.M., Johnston D.T., Hansel C.M., Joye S.B., Girguis P.R., Anaerobic methane oxidation in metalliferous hydrothermal sediments: influence on carbon flux and decoupling from sulfate reduction, *Environ. Microbiol.*, 14, 2012, 2726–2740.

- [62] Welte C.U., Nitrate- and nitrite-dependent anaerobic oxidation of methane, *Environ. Microbiol. Rep.*, 8, 2016, 941–955.
- [63] Zehnder A.J., Brock T.D., Methane formation and methane oxidation by methanogenic bacteria, *J Bacteriol.*, 137(1), 1979, 420–432.

*Przesłano do redakcji: 18.03.2019 r.*